

Immunopathological Concepts Behind Autoimmunity

SOMA SUSAN VARGHESE¹, SREENIVASAN BARGAVAN SAROJINI², PHILIPS MATHEW³, JAYAN JACOB MATHEW⁴

ABSTRACT

Autoimmunity is the failure of self-tolerance of an organism, which empowers immune response against its own cells and tissues generating autoimmune diseases. Autoimmune diseases are exceeding 100 million people globally. Autoimmune mechanisms underline many diseases, some are organ-specific and others are systemic in distribution. Aberrations in central and peripheral tolerance, molecular mimicry, bystander activation, cryptic antigenic and super-antigenic exposure are found to be associated with immune response against self-antigens. As the molecular biology behind autoimmune diseases is obscure, an insight on immune pathways, impaired immunity and evolution of autoimmune diseases can supplement on the therapeutic modalities of immunological diseases.

Keywords: Autoimmunity, Cryptic antigen, Epitope spreading, Molecular mimicry, Self tolerance

INTRODUCTION

Immune system refers to the assortment of defence cells against foreign antigens. The line of defence in the immune system encompasses Innate and Adaptive immune responses [1]. D2-Immune system is under the control and regulation of inflammatory cells which safeguard and prevent body from cellular injury and infections. Inflammatory cells can also act as a “double-edged sword” in both protective and destructive manner emphasizing the role of immune cells in autoimmune mechanism wherein self antigens are recognised as antigenic. Instant recognition of encroaching foreign antigen is perceived by innate immune mechanism which is deprived of immunological memory. Adaptive immune response is antigen dependent and specific with immunologic memory which empowers immune mechanism upon succeeding antigenic response [2]. Central intra-thymic deletion of self-reactive T lymphocytic cells which own receptors for self-antigen is self-tolerance [3]. Inhibition of self-antigen reactive immune cells in the periphery is performed by regulatory T-cells (T regs) [4,5]. Some T cells termed as ignorant T-cells are neither deleted in the periphery nor in the thymus due to the diminished amount of their cryptic antigen [6]. D3-Antigen Presenting Cells (APC) along with surface Major histocompatibility complex present antigen to T-lymphocytes. CD4+T cells are restricted by MHC class II antigens and CD8+ cells are restricted by MHC class I antigens. Major histocompatibility (MHC) complex comprising HLA (Human Leukocyte Antigen) class I and HLA class II could perceive foreign antigens and present to T-lymphocytic receptors [7]. Self Non-Self (SNS) model, Infection Non-Self (INS) model, Danger model and Two Signal model are considered as some of the ideal concepts of immune reactions [8]. Autoimmune disease is the sequelae of impairment of self-tolerance to self-antigen.

Autoimmune diseases are exceeding 100 million people globally [9]. Cessation of central and peripheral tolerance is found to be associated with prolonged environmental insults generating immune response against self-antigens. Molecular mimicry, bystander activation, cryptic antigenic and super antigenic exposure can promote autoimmune reactions [10-12].

Concepts in Autoimmunity

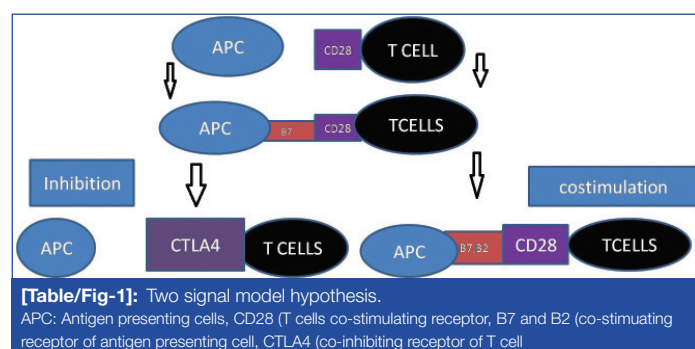
Self Non-Self model theory and Two Signal model theory: The SNS model theory of immunity states that any non-self or foreign

antigen can trigger immune mechanism in the host, whereas self endogenous elements will not provoke an immune reaction in the host. Confrontation of body with any foreign material can bring about immune response. A contrast statement was proposed by Jerne which recommends that a constant negligible reaction should be there by immune cells on self molecules for normal immune surveillance [13]. During lymphocytic selection in the thymus or bone marrow, these immune cells could survive in the primary lymphoid organs only if it reacts on self molecule. Deprivation of weak reaction of circulating lymphocyte on self antigen can result in its cessation. T regulatory lymphocytes which are self immune cells control the normal self lymphocytes during immune functions [14-16]. Immune tolerance of host immune cells to gut bacteria, certain helminths, alloantigenic graft, chimerism and foetomaternal tolerance are countering SNS model theory of immunology [17-20].

In the initial phase of SNS model, it was postulated that interaction of B-cell antigenic receptors and T-cell antigenic receptors with an antigen can generate immune reactions. Subsequently two signal model theory of immunology was implied, declaring that B-cell after processing antigen, re-express it on MHC class II molecule for T-helper cells to identify [21,22]. A consequent theorization was proposed recommending that T-lymphocytes need stimulatory signals from Antigen Presenting Cells (APC) for immune reaction. TCR signalling is monitored by co-stimulatory and co-inhibitory receptors on its surface. After the recognition of antigenic peptides presented by MHC molecules to T-Cell Receptors (TCR), T-cell function and fate is determined by the co-stimulatory and co-inhibitory receptors on T-cells.

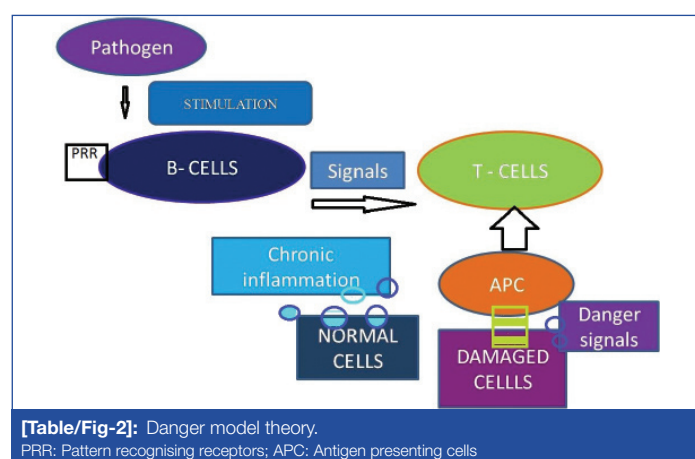
According to the two signal hypothesis of immunity, both antigen and secondary stimuli are required for T cell activation. The most complex two signal model, immune regulatory system operates with the conjugation of the co-stimulatory receptor CD28 (ligand, B7-1), and co inhibitory receptor (Cytotoxic T Lymphocyte Antigen-4 (CTLA4), which also binds to B7-1) and a second ligand (B7-2, which binds to both CD28 and CTLA4) [23-25]. Expression of B7-1 and B7-2 is modulated by the activation state of the APC. T cell growth and survival is promoted by CD28 co stimulatory receptor, expressing on the cell surface of naive CD4+ and CD8+T cells upon ligation by B7-1 and B7-2 on antigen-presenting cells (APCs) [26-28]. Instigation of CTLA4, co inhibitory receptor is promoted by the activation of T cell and increased expression of CTLA4 declines

CD28 expression by endocytosis [29-31]. Cellular insults activate APCs and induce transcription, translation and transportation of both B7-1 and B7-2 to the cell surface [32,33]. The formation of the immune synapse allows TCR signalling and co-signalling [34,35] [Table/Fig-1] D1. The central, peripheral and distal Supra-Molecular Activation Complexes (cSMAC, pSMAC and dSMAC) form the immune synapse with TCR which provides evidence for the two-signal model of T cell activation [36,37]. Co signalling ligands and



counter-receptors are highly expressed on the professional Antigen Presenting Cells (APCs).

Danger model theory: Danger model hypothesis was postulated by Polly Matzinger, according to which the immune system is more concerned with safe guarding the individual from insults rather than discriminating self and non-self. Circumstances like Mechanical stress, DNA damage, heat, cold and hypoxia can result in cellular damage. Injured, distressed, damaged or necrotic cells could liberate danger or alarm signals which stimulate APC whereas healthy cells send calm signals and apoptotic cells convey eat me signals to APC [38-40]. Repeated cellular insults generate danger alarm signals augmenting antigen presentation which in due course recognises, self protein antigenic, precipitating chronic autoimmunity D2 [Table/Fig-2] [41,42]. A S8-Some of the Endogenous signals are



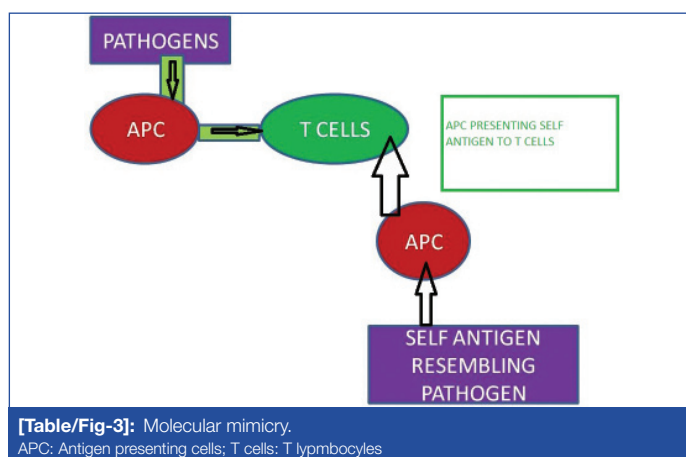
Heat Shock Proteins (HSP), hyaluronan, High Mobility Group Box-1 Mediator (HMGB1), Fibronectin fragments, modified low density lipoprotein and extracellular ATP [43-48].

HSP which are released on account of cellular stress could switch on innate immune cell via Toll-Like Receptors (TLRs). HSP induces the production of nitric oxide (NO), down regulation of tumour necrosis factor α , activates IL12 and regulatory T-cells and improves antigen presentation if associated with antigen [49-51]. The sulphated glycosaminoglycans, hyaluronan which is a major component of extracellular matrix is cleaved to hyaluronic acid during tissue injury [52,53]. This signalling response could institute innate immune response and propagation of dendritic cell maturation contributing adaptive immune response. HMGB1 is released from necrosed cells with the aid of macrophages and dendritic cells. HMGB1 with

its endogenous stimulatory mechanism can contribute autoimmune reaction with existing chronic inflammation [54,55]. Fibronectin fragments, defensin and modified low density lipoprotein through divergent immune response can contribute to immune reaction.

Immunopathology of Autoimmune Reactions

Molecular Mimicry: Elicitation of immune response against self antigens, when there exist an antigenic resemblance between foreign peptide or pathogen and self peptide is called molecular mimicry [56]. The central and peripheral tolerance which is specifically the positive and negative selection of primitive T cells occurs in the thymus [57]. The selection depends on the competence of T cells to interact with the peptides presented by MHC. T-cells, with negative selection of MHC presented peptides and strong interactions with self antigens are nullified [58,59]. Instead in molecular mimicry a



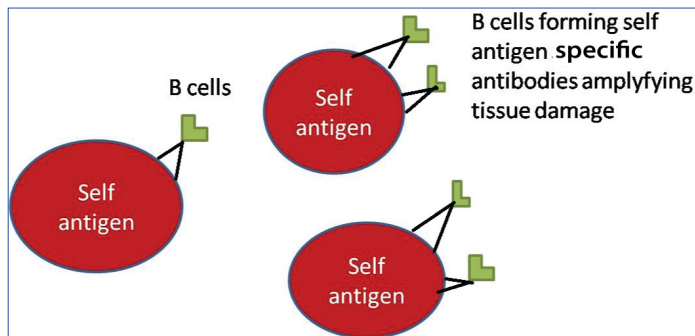
constant immune response against self-antigens is ensued, when the pathogen has a structural and sequential resemblance with self antigens [Table/Fig-3]. Molecular mimicry can also be emerged by virtue of neopeptide formation acquiring via tissue injury. Tissue injury can contribute to the development of self reactive T cells by exposing the self cryptic epitopes [60,61].

AS10-TCR is poly specific in nature. The structural resemblance of pathogens to self antigens creates dilemma to TCR inducing molecular mimicry [62]. A foreign peptide (such as virus) resembling MHC derived peptide can activate T cells, and if the self antigen is also structurally similar, exposure to the foreign peptide results in these activated T cell becoming autoreactive [63,64]. Molecular mimicry is confederated to several autoimmune diseases like Multiple sclerosis, Gullian-Barre syndrome, Acute Rheumatic fever, type 1 Diabetes Mellitus, Lyme arthritis and Ankylosing Spondylitis. A structural homology between Epstein-Barr virus and Myelin basic protein was evident in multiple sclerosis [65]. In Gullian-Barre syndrome post infectious molecular mimicry between nerve antigen and microbes induce autoimmune response [66]. Epitope mimicry between Streptococcal M proteins and human myosin, tropomyosin, keratin, actin, laminin, vimentin and N-acetylglucosamine contribute to acute rheumatic fever [67]. In type 1 diabetes mellitus, mimicry related to viral infection is hypothesized due to the structural similarity between pancreatic beta cell enzyme and viruses [68,69].

Bystander Activation: In bystander activation, viral and bacterial products induce INF α/β (Interferon alfa/beta) secretion which propagates the proliferation and expansion of heterologous polyclonal T cells [70]. Activation of INF α/β D11 (Interferon alfa / beta) leads to the stimulation of APC [71]. APC produce the cytokine, IL -15 which then activates memory phenotype (MP CD+8) T cells. CD8+T cells recognize the infected cells and release cytokines like TNF (Tumour Necrosis Factor), NO (Nitric Oxide) and Lymphocyte Toxins (LT) resulting in the causation of bystander killing of uninfected neighbouring cells D3 [72]. Cytokines released from CD4+T cells can directly kill uninfected cell but the bystander

activation of CD4+T cells are less competent than CD8+T Cells. CD 122 receptor for IL2/IL15 is more expressed by MP CD8+T cells than MP CD4+T which results in limited bystander activation of MP CD4+T [73].

Epitope Spreading: AS13-Epitope spreading can be defined as a specific autoreactive lymphocyte (T or B cell) response to endogenous epitopes, secondary to the release of such self protein during a chronic autoimmune or inflammatory response (Vanderlugt and Miller, 1996) [Table/Fig-4] [74]. Cell mediated and humoral immunities are influenced in epitope spreading by the unmasking of multiple epitopes on a single antigen or from one antigenic molecule to other. The biological events elucidated in epitope spreading



[Table/Fig-4]: Epitope spreading showing clonal expansion of autoantibodies causing damage on self antigens.

include endocytic processing, antigen presentation, and somatic hypermutation. B cell epitope spreading appeared to be inveigled in autoimmune diseases like Systemic lupus erythematosus, multiple sclerosis, pemphigus and bullous pemphigoid [75]. T cell epitope spreading is accomplished in Multiple Sclerosis and Autoimmune Encephalomyelitis, by a concurrent regression of primary autoreactivity associated with disease onset [76].

CONCLUSION

Insight on immune pathways, impaired immunity and evolution of autoimmune diseases can supplement on the therapeutic modalities of immunological diseases. Distraction in the suppression of the immune response to the self antigens contributes to autoimmune diseases. Various mechanisms, including the release of sequestered antigens, molecular mimicry, epitope spreading, cytokine deregulation and inappropriate expression of class II MHC molecules propagate autoimmunity.

REFERENCES

- [1] Turvey SE, Broide DH. Innate immunity. *J Allergy Clin Immunol*. 2010;125(2):24-32.
- [2] Sun JC, Ugolini S, Vivier E. Immunological memory within the innate immune system. *EMBO J*. 2014;33(12):1295-303.
- [3] Griesemer AD, Sorenson EC, Hardy MA. Role of thymus in tolerance. *Transplantation*. 2010;90(5):465-74.
- [4] Sakaguchi S, Wing K, Miyara M. Regulatory T cells—a brief history and perspective. *Eur J Immunol*. 2007;37:116-23.
- [5] Sojka DK, Huang YH, Fowell DJ. Mechanisms of regulatory T-cell suppression—a diverse arsenal for a moving target. *Immunology*. 2008;124:13-22.
- [6] Kurts C, Sutherland RM, Davey G, Li M, Lew AM, Blanas E. CD8 T cell ignorance or tolerance to islet antigens depends on antigen dose. *Proc Natl Acad Sci U S A*. 1999;96(22):12703-07.
- [7] Miles JJ, McCluskey J, Rossjohn J, Gras S. Understanding the complexity and malleability of T-cell recognition. *Immunology and Cell Biology*. 2015;93:433-41.
- [8] Matzinger P. The danger model: a renewed sense of self. *Science*. 2002;296:301-05.
- [9] Cooper GS, Bynum ML, Somers EC. Recent insights in the epidemiology of autoimmune diseases: improved prevalence estimates and understanding of clustering of diseases. *J Autoimmun*. 2009;33(3-4):197-207.
- [10] Fujinami RS, Oldstone MBA. Amino acid homology between the encephalitogenic site of myelin basic protein and virus: Mechanism for autoimmunity. *Science*. 1985;230:1043-45.
- [11] Oldstone MBA. Molecular mimicry and autoimmune disease. *Cell*. 1987;50:819-20.
- [12] McRae BL, Vanderlugt CL, Dal Canto MC, Miller SD. Functional evidence for epitope spreading in the relapsing pathology of experimental autoimmune encephalomyelitis. *J Exp Med*. 1995;182:75-85.

- [13] Pradeu T, Carocella ED. The self model and the conception of biological identity in immunology. *Biology and Philosophy*. 2006;21:235-52.
- [14] Loh DY, Sha WC, Nelson CA, Newberry RD, Kranz, Russell JH. Positive and negative selection of T lymphocytes. *Cold Spring Harb Symp Quant Biol*. 1989;54:147-51.
- [15] Goodnow CC. Balancing immunity and tolerance: deleting and tuning lymphocyte reperpetoires. *Proc Natl Acad Sci USA*. 1996;93(6):2264-71.
- [16] Hoynes GF, Lamb JR. Regulation of T cell function in mucosal tolerance. *Immunology and Cell Biology*. 1997;75:197-201.
- [17] Berg RD. The indigenous gastrointestinal microflora. *Trends in Microbiology*. 1996;4:430-35.
- [18] Buscaglia CA, Di Noia JM. *Trypanosoma cruzi* clonal diversity and the epidemiology of Chagas' disease. *Microbes Infect*. 2003;5(5):419-27.
- [19] Ferguson TA, Griffith TS. A vision of cell death: insights into immune privilege. *Immunological review*. 1997;156:167-84.
- [20] Aluvihare VR. Regulatory T cells mediate maternal tolerance to the fetus. *Nat Immunol*. 2004;5(3):266-71.
- [21] Langman RE, Cohn M. Self-non-self discrimination revisited. *Semin Immunol*. 2000;12:159-62.
- [22] Cohn M, Langman RE. The protection: the unit of humoral immunity selected by evolution. *Immunol Rev*. 1990;115:11-147.
- [23] Linsley PS. CTLA-4 is a second receptor for the B cell activation antigen B7. *J Exp Med*. 1991;174:561-69.
- [24] Azuma M. B70 antigen is a second ligand for CTLA-4 and CD28. *Nature*. 1993;366:76-79.
- [25] Hathcock KS. Identification of an alternative CTLA-4 ligand costimulatory for T-cell activation. *Science*. 1993;262:905-07.
- [26] Knieke K, Lingel H, Chamaon K, Brunner-Weinzierl MC. Migration of Th1 lymphocytes is regulated by CD152 (CTLA-4)-mediated signaling via PI3 kinase-dependent Akt activation. *Plos One*. 2012;7:1-9.
- [27] Hathcock KS, Laszlo G, Pucillo C, Linsley P, Hodes RJ. Comparative analysis of B7-1 and B7-2 costimulatory ligands: expression and function. *J Exp Med*. 1994;180:631-40.
- [28] Boomer JS, Green JM. An enigmatic tail of CD28 signaling. *Cold Spring Harb Perspect Biol*. 2010;2:01-12.
- [29] Janardhan SV, Praveen K, Marks R, Gajewski TF. Evidence implicating the Ras pathway in multiple CD28 costimulatory functions in CD4+T cells. *Plos One*. 2011;6:01-10.
- [30] Saito T, Yokosuka T, Hashimoto-Tane A. Dynamic regulation of T cell activation and co-stimulation through TCR-microclusters. *FEBS Lett*. 2010;584:4865-71.
- [31] Koshland DE. Recognising self from non self. *Science*. 1990;248:1273.
- [32] Matzinger P. Tolerance, danger, and the extended family. *Annu Rev Immunol*. 1994;12:991.
- [33] Rock KL, Kono H. The inflammatory response to cell death. *Annu Rev Pathol*. 2008;3:99-126.
- [34] Davis DM, Dustin ML. What is the importance of the immunological synapse? *Trends Immunol*. 2004;25:323-27.
- [35] Thauland TJ, Parker DC. Diversity in immunological synapse structure. *Immunology*. 2010;131:466-72.
- [36] Dustin ML, Chakraborty AK, Shaw AS. Understanding the structure and function of the immunological synapse. *Cold Spring Harb Perspect*. 2010;2(10):002311.
- [37] Griffiths GM, Tsun A, Stinchcombe JC. The immunological synapse: a focal point for endocytosis and exocytosis. *J Cell Biol*. 2010;189:399-406.
- [38] Matzinger P. Friendly and dangerous signals: is the tissue in control? *Nat Immunol*. 2007;8:11-13.
- [39] Matzinger P. The evolution of the danger theory. *Expert Rev Clin Immunol*. 2012;8:311-17.
- [40] Matzinger P, Kamala T. Tissue-based class control: the other side of tolerance. *Nat Rev Immunol*. 2011;11:221-30.
- [41] Akira S, Uematsu S, Takeuchi O. Pathogen recognition and innate immunity. *Cell*. 2006;124:783-801.
- [42] Akira S, Takeda K. Toll-like receptor signalling. *Nat Rev Immunol*. 2004;4:499-511.
- [43] Kaufmann SH. Heat shock proteins and the immune response. *Immunol Today*. 1990;11:129.
- [44] Van Eden W, Van der Zee R, Paul AG, Prakken BJ, Wendling U, Anderton SM, et al. Do heat shock proteins control the balance of T-cell regulation in inflammatory diseases? *Immunol Today*. 1998;19:303.
- [45] Scheibner KA. Hyaluronan fragments act as an endogenous danger signal by engaging TLR2. *J Immunol*. 2006;177:1272-81.
- [46] Kovacs-Simon A, Titball RW, Michell SL. Lipoproteins of bacterial pathogens. *Infect Immun*. 2011;79(2):548-61.
- [47] Vohra RS, Murphy JE, Walker JH, Ponnambalam S, Homer-Vanniasinkam S. Atherosclerosis and the Lectin-like Oxidized low-density lipoprotein scavenger receptor. *Trends Cardiovasc Med*. 2006;16:60-64.
- [48] Miller YI, Viriyakosol S, Binder CJ, Feramisco JR, Kirkland TN, Witztum JL. Minimally modified LDL binds to CD14, induces macrophage spreading via TLR4/MD-2, and inhibits phagocytosis of apoptotic cells. *J Biol Chem*. 2003;278:1561-68.
- [49] Ohshima H, Bartsch H. Chronic infections and inflammatory processes as cancer risk factors: possible role of nitric oxide in carcinogenesis. *Mutat Res*. 1994;305:253-64.
- [50] Huang QQ, Sobkoviak R, Jockheck-Clark AR, Shi B, Mandelin AM, Tak PP, et al. Heat shock protein 96 is elevated in rheumatoid arthritis and activates macrophages primarily via TLR2 signaling. *J Immunol*. 2009;182:4965-73.
- [51] Singh IS, He JR, Calderwood S, Hasday JD. A high affinity HSF-1 binding site in the 5'-untranslated region of the murine tumor necrosis factor-alpha gene is a transcriptional repressor. *J Biol Chem*. 2002;277(7):4981-88.
- [52] Termeer C, Benedix F, Sleeman J, Fieber C, Voith U, Ahrens T, et al. Oligosaccharides of hyaluronan activate dendritic cells via toll-like receptor 4. *J*

- Exp Med. 2002;195:99-111.
- [53] Scheibner KA, Lutz MA, Boodoo S, Fenton MJ, Powell JD, Horton MR. Hyaluronan fragments act as an endogenous danger signal by engaging TLR2. *J Immunol.* 2006;177:1272.
- [54] Abraham E, Arcaroli J, Carmody A, Wang H, Tracey KJ. HMG-1 as a mediator of acute lung inflammation. *J Immunol.* 2000;165:2950-54.
- [55] Lotze MT, Tracey KJ. High-mobility group box 1 protein (HMGB1): nuclear weapon in the immune arsenal. *Nat Rev Immunol.* 2005;5:331-42.
- [56] Cusick MF, Libbey JE, Fujinami RS. Molecular mimicry as a mechanism of autoimmune disease. *Clin Rev Allergy Immunol.* 2012;42(1):102-11.
- [57] Egerton M, Scollay R, Shortman K. Kinetics of mature T-cell development in the thymus. *Proc Natl Acad Sci USA.* 1990;87(7):2579-82.
- [58] Starr TK, Jameson SC, Hogquist KA. Positive and negative selection of T cells. *Annu Rev Immunol.* 2003;21:139.
- [59] Takahama Y. Journey through the thymus: stromal guides for T-cell development and selection. *Nat Rev Immunol.* 2006;6(2):127.
- [60] Powell AM, Black MM. Epitope spreading: protection from pathogens, but propagation of autoimmunity? *Clin Exp Dermatol.* 2001;26:427-33.
- [61] Bonsor DA, Grishkovskaya I, Dodson EJ, Kleanthous C. Molecular mimicry enables competitive recruitment by a natively disordered protein. *J Am Chem Soc.* 2007;129: 4800-07.
- [62] Lang HL, Jacobsen H, Ikemizu S, Andersson C, Harlos K, Madsen L, et al. A functional and structural basis for TCR cross-reactivity in multiple sclerosis. *Nat Immunol.* 2002;3(10):940-43.
- [63] Wucherpfennig KW, Strominger JL. Molecular mimicry in T cell-mediated autoimmunity: viral peptides activate human T cell clones specific for myelin basic protein. *Cell.* 1995;80:695-705.
- [64] Evavold BD, Sloan-Lancaster J, Wilson KJ, Rothbard JB, Allen PM. Specific T cell recognition of minimally homologous peptides: evidence for multiple endogenous ligands. *Immunity.* 1995;2:655-63.
- [65] Libby JE, MC Coy LL, Fujinami RS. Molecular mimicry in multiple sclerosis. *Int Rev Neurobiol.* 2007;79:127-47.
- [66] Ang CW, Jacobs BC, Laman JD. The Guillain-Barré syndrome: a true case of molecular mimicry. *Trends Immunol.* 2004;25:61-66.
- [67] Guilherme L, Ramasawmy R, Kallil J. Rheumatic fever and rheumatic heart disease: genetics and pathogenesis. *Scandinavian Journal of Immunology.* 2007;66:199-207.
- [68] Fujinami RS, Oldstone MB. Molecular mimicry as a mechanism for virus-induced autoimmunity. *Immunol Res.* 1989;8:3.
- [69] Benoist C, Mathis D. Autoimmunity provoked by infection: how good is the case for T cell epitope mimicry? *Nat Immunol.* 2001;2:797-801.
- [70] Boyman O. Bystander activation of CD41 T cells. *Eur J Immunol.* 2010;40:936-39.
- [71] Tough DF, Zhang X, Sprent J. An IFN-gamma-dependent pathway controls stimulation of memory phenotype CD81 T cell turnover in vivo by IL-12, IL-18, and IFN-gamma. *J Immunol.* 2001;166:6007-11.
- [72] Eberl G, Brawand P, MacDonald HR. Selective bystander proliferation of memory CD41 and CD81 T cells upon NK T or T cell activation. *J Immunol.* 2000;165:4305-11.
- [73] Berard M, Thoug DF. Qualitative differences between naïve and memory T cells. *Immunology.* 2002;106(2):127-38.
- [74] Trentham DE, Townes AS, Kang AH. Cellular sensitivity to collagen in rheumatoid arthritis. *Archivum Immunologiae et Therapiae Experimentalis.* 2000;8:347-51.
- [75] Cornbay C, Gibbons L, Meyhew V, Sloan CS. B cell epitope spreading: Mechanisms and contribution to autoimmune diseases. *Immunology Letters.* 2015;163:56-68.
- [76] Tuohy VK, Yu M, Yin L, Kawczak JA, Johnson JM, Mathisen PM, et al. The epitope spreading cascade during progression of experimental autoimmune encephalomyelitis and multiple sclerosis. *Immunol Rev.* 1998;164:93-100.

PARTICULARS OF CONTRIBUTORS:

1. Reader, Department of Oral Pathology and Microbiology, Marbaselois Dental College, Kothamangalam, Ernakulam, Kerala, India.
2. Professor, Department of Oral Pathology and Microbiology, Marbaselois Dental College, Kothamangalam, Ernakulam, Kerala, India.
3. Assistant Professor, Department of Oral Medicine and Radiology, Government Dental College, Kottayam, Kerala, India.
4. Professor, Department of Periodontics, Marbaselois Dental College, Kothamangalam, Ernakulam, Kerala, India.

NAME, ADDRESS, E-MAIL ID OF THE CORRESPONDING AUTHOR:

Soma Susan Varghese,
Reader, Department of Oral Pathology and Microbiology, Marbaselois Dental College, Kothamangalam, Ernakulam, Kerala, India.
E-mail: drsomasusan@yahoo.in

Date of Submission: **Mar 01, 2018**
Date of Peer Review: **Apr 28, 2018**
Date of Acceptance: **Jun 25, 2018**
Date of Publishing: **Oct 01, 2018**

FINANCIAL OR OTHER COMPETING INTERESTS: None.